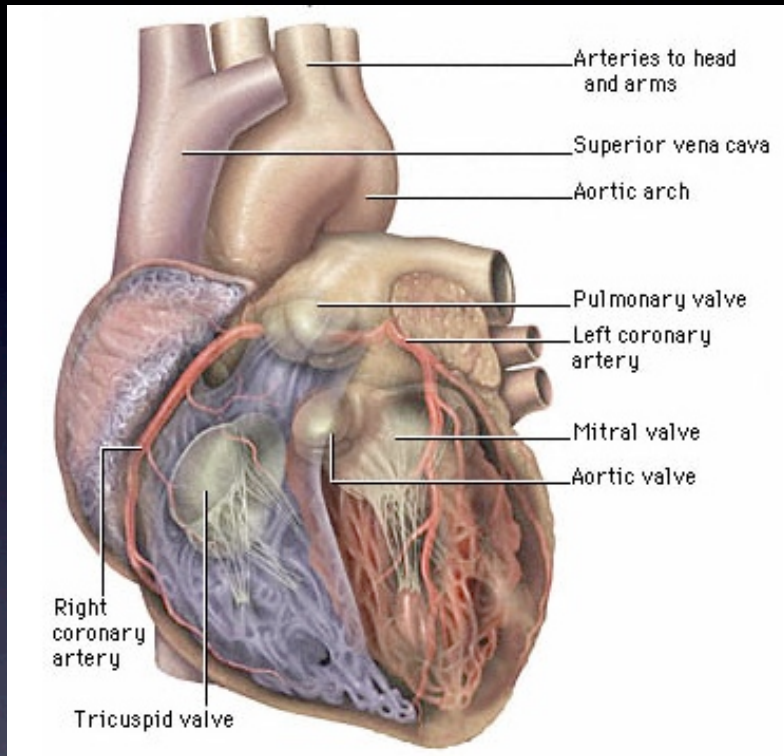


# What Can We Learn From Integrative Modeling of the Heart ?

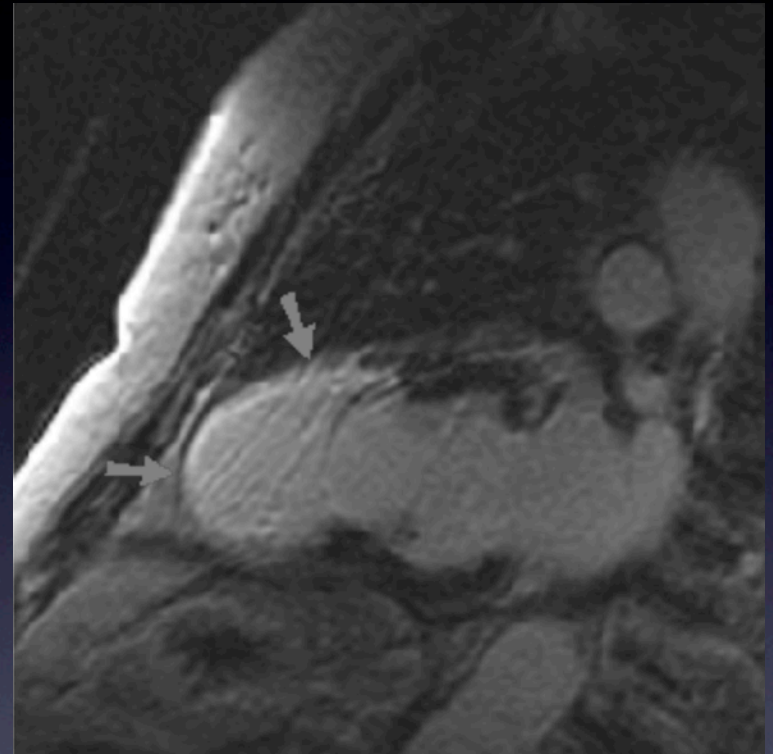
Raimond L. Winslow

The Institute for Computational Medicine &  
Department of Biomedical Engineering  
The Johns Hopkins University

# The Human Heart in Health and Disease



~ 3 billion beats per lifetime



Courtesy Dr. Joao Lima

# Vertical Integration

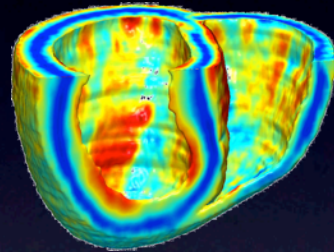
**Domain**

**Biological Hierarchy**

**Spatial Scale**

**Time Scale**

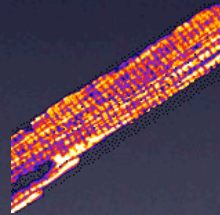
Heart (Macro-scale)



$\sim 10^5 \mu\text{m}$

$\sim 10^2 \text{ Sec}$

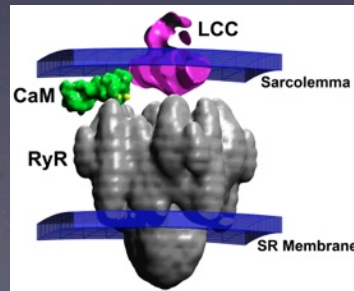
Cells (Meso-scale)



$\sim 10^2 \mu\text{m}$

$\sim 10^{-3} \text{ Sec}$

Molecules (Nano-scale)



$\sim 10^{-3} \mu\text{m}$

$\sim 10^{-9} \text{ Sec}$

Physiology



Cell  
Biology



Molecular  
Biology

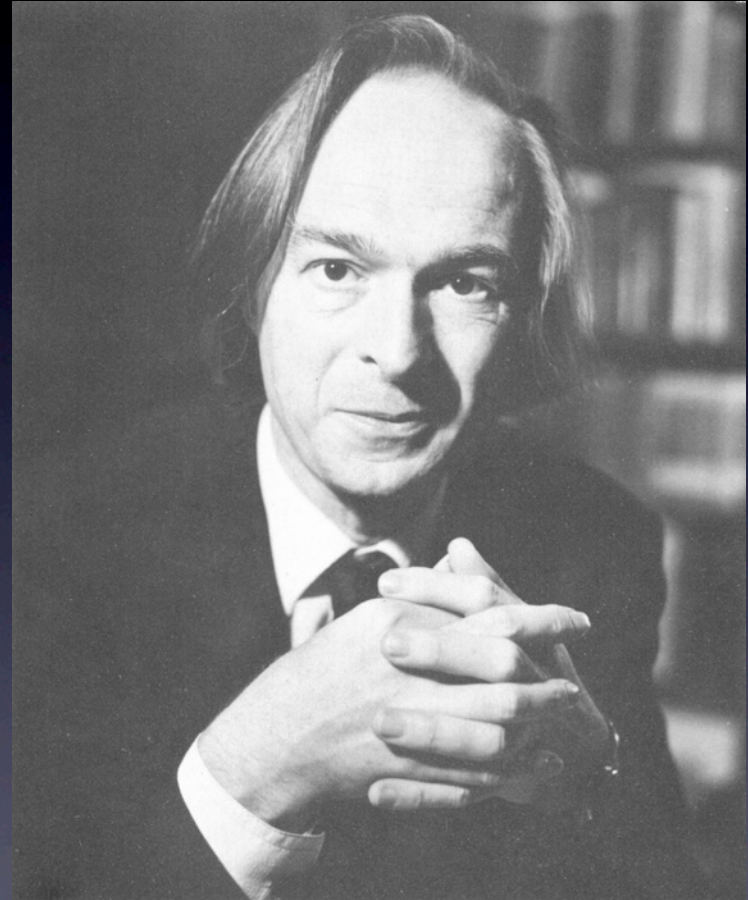
# 50 Years of Cardiac Modeling !

## Cardiac Action and Pacemaker Potentials based on the Hodgkin-Huxley Equations

SINCE the equations describing the nerve action potential were formulated by Hodgkin and Huxley<sup>1</sup>, the range of phenomena to which they have been shown to apply has been greatly extended. Huxley<sup>2</sup> has applied them to the influence of temperature on the propagated response and to the repetitive firing observed in low calcium concentrations. More recently, Fitzhugh<sup>3</sup> has shown that the long action potentials induced by tetraethylammonium ions in squid nerve may also be reproduced.

The computations described in this communication were carried out with the aim of reconstructing the long-lasting action potential and pacemaker potential of cardiac muscle. Although this work was done independently, the results agree with those of Fitzhugh in showing that action potentials of long duration may be accounted for by Hodgkin and Huxley's formulation of the membrane properties. The description of the potassium current, however, differs from that used by Fitzhugh and provides a better description of the conductance changes.

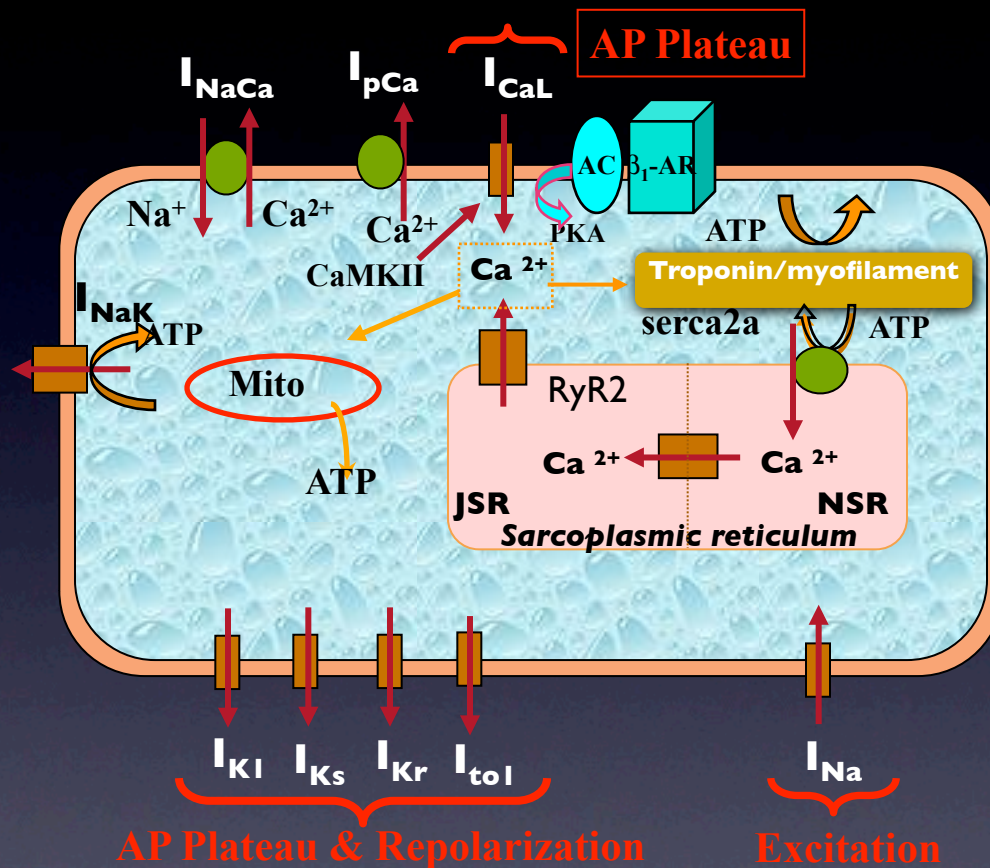
The equations I have used to describe the sodium current are very similar to Hodgkin and Huxley's,



D. Noble (1960). *Nature*, 188:495  
November 5, 1960

Denis, *circa* 1984

# Integrative Cell Models: State of the Art



## • Constraining data

- Functional measurements (t)
- *low-throughput*

## • Model Components

- Ion channels & membrane transporters
- $Ca^{2+}$ -induced  $Ca^{2+}$ -release (CICR) and  $Ca^{2+}$  cycling
- $Ca^{2+}$ -driven force generation
- Mitochondrial energetics
- CaMKII, PKA signaling

Top

Down

# Integrative Modeling Case Study

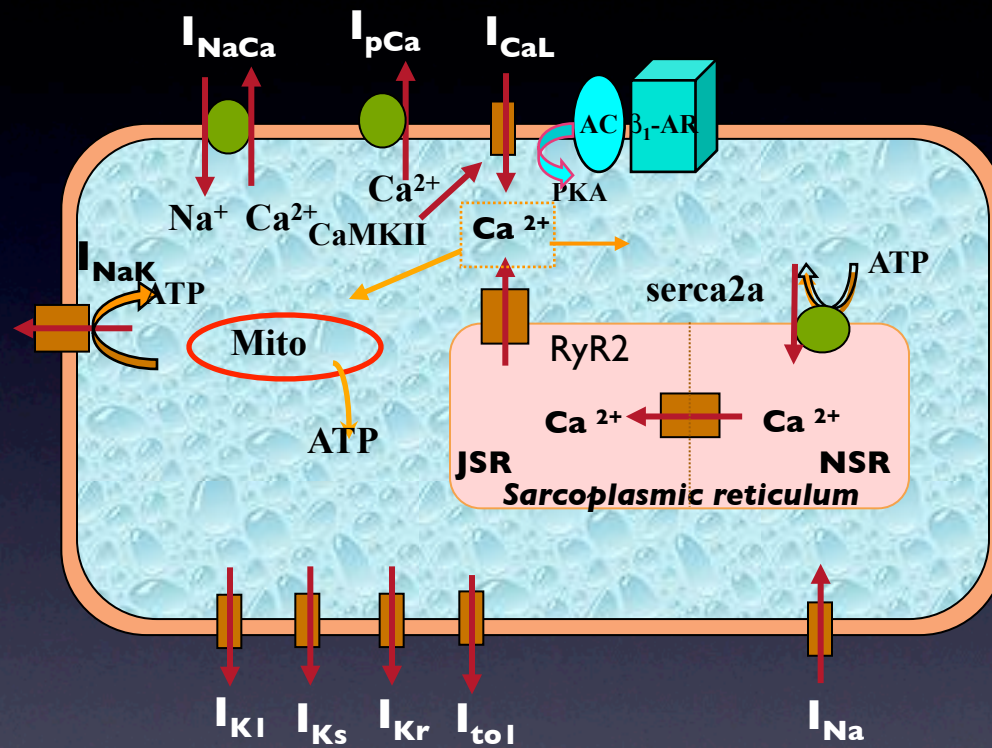
## Heart Failure

- Diverse origins, common phenotype
- Reduced contractility, mechanical pump failure
- Increased risk of Sudden Cardiac Death
- *The* primary U.S. hospital discharge diagnosis
  - Incidence ~ 400,000/year, prevalence ~ 4.5 million
  - 15% mortality at 1 Yr, 80% mortality at 6 Yr
  - Medical expenditures ~ \$20 billion per year

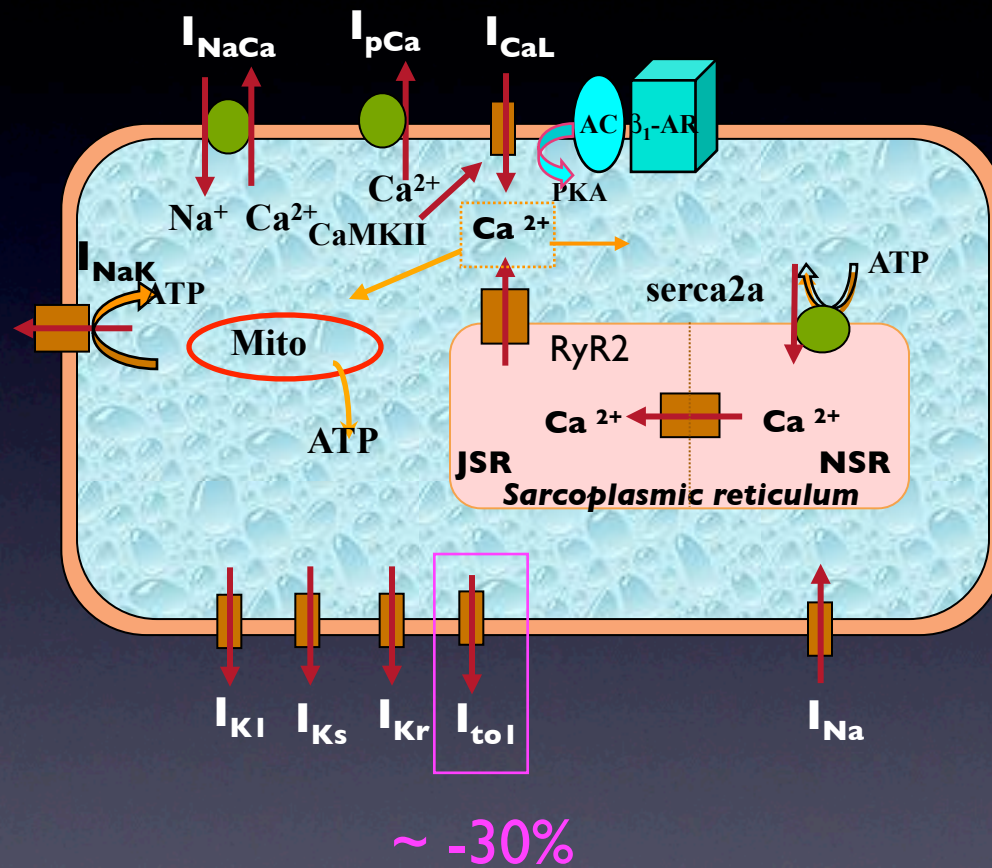
# Using Models to Understand the Phenotypes of Heart Failure

- ***Molecular phenotype***
  - altered protein expression
  - disruption of protein co-localization
- ***Pathway phenotype***
  - increased B-adrenergic input, PKA hyper-phosphorylation
- ***Cellular phenotype***
  - Action potential prolongation, repolarization abnormalities
- ***Organ phenotype***
  - Reduced contractility
  - High risk of re-entrant arrhythmias

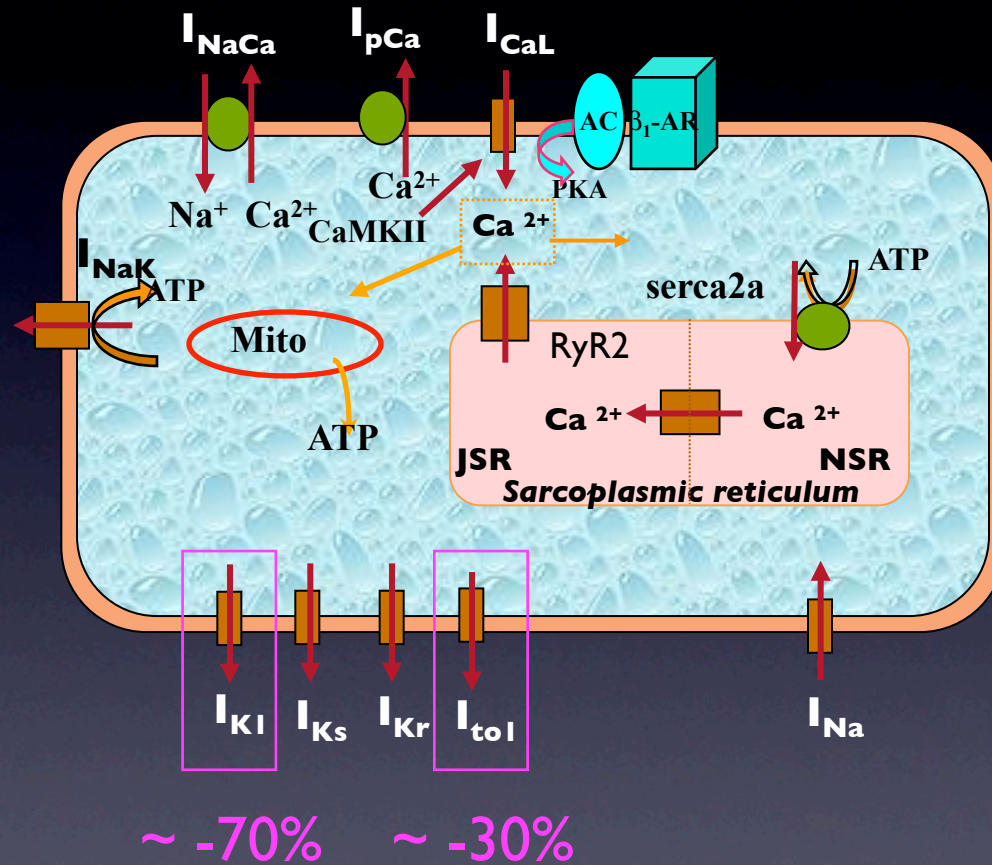
# The *Molecular Phenotype* of Heart Failure



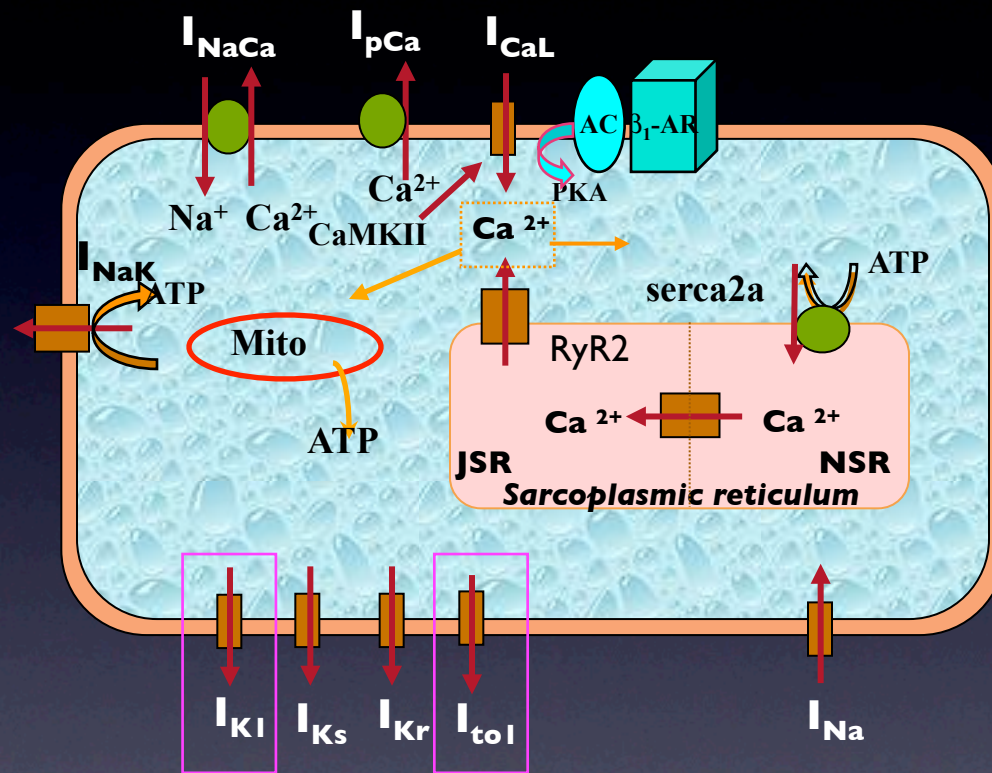
# The Molecular Phenotype of Heart Failure



# The Molecular Phenotype of Heart Failure



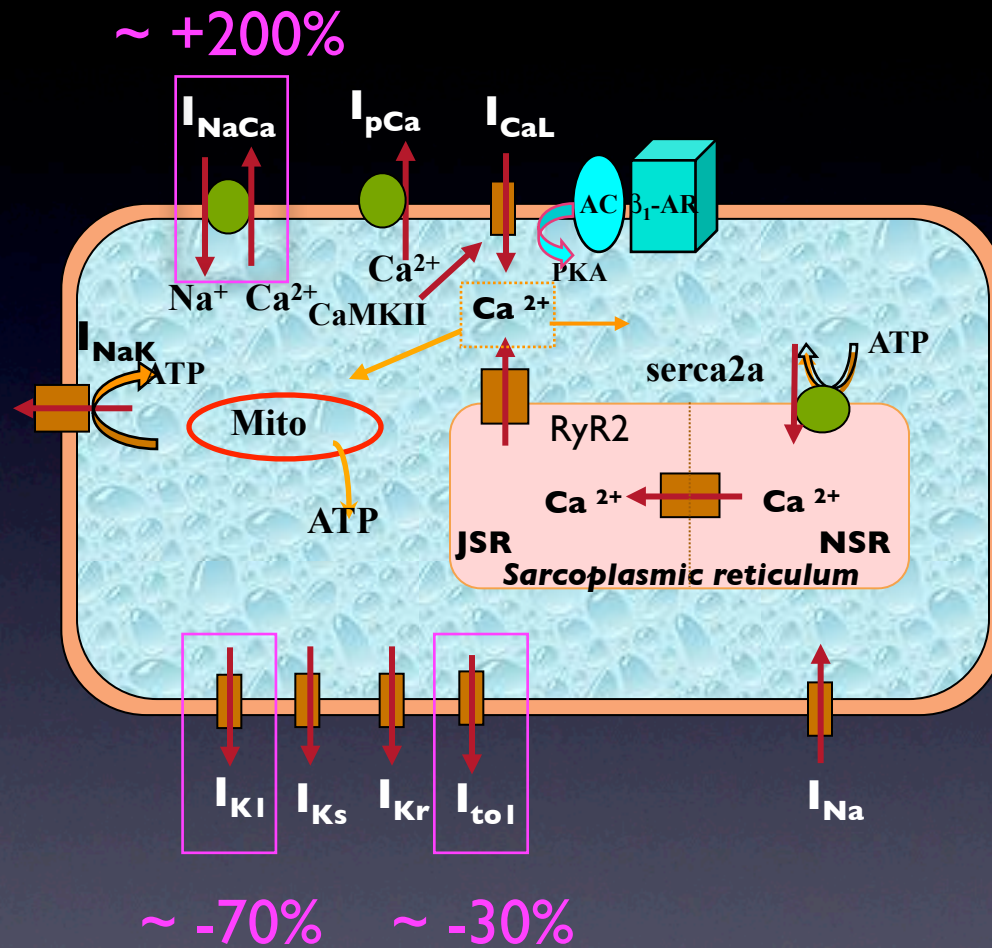
# The *Molecular Phenotype* of Heart Failure



~ -70%    ~ -30%

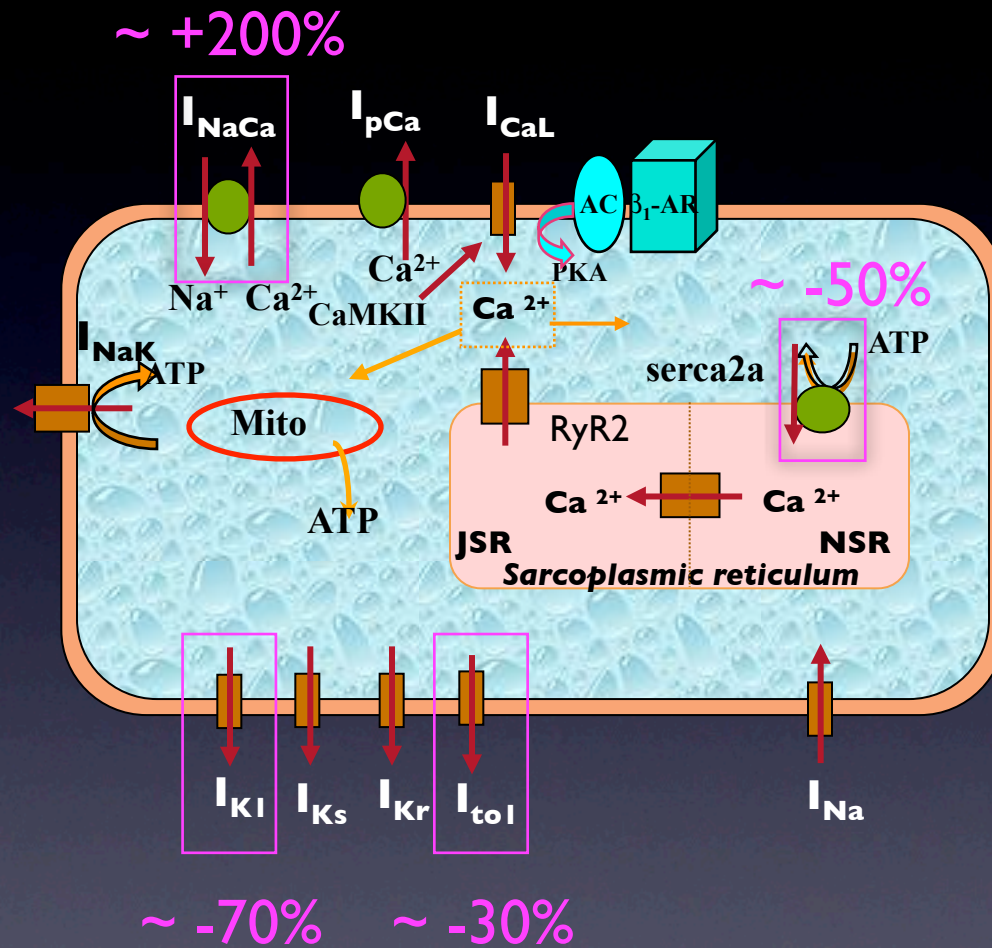
Hypothesis: Reduced Outward Current Prolongs APD

# The Molecular Phenotype of Heart Failure



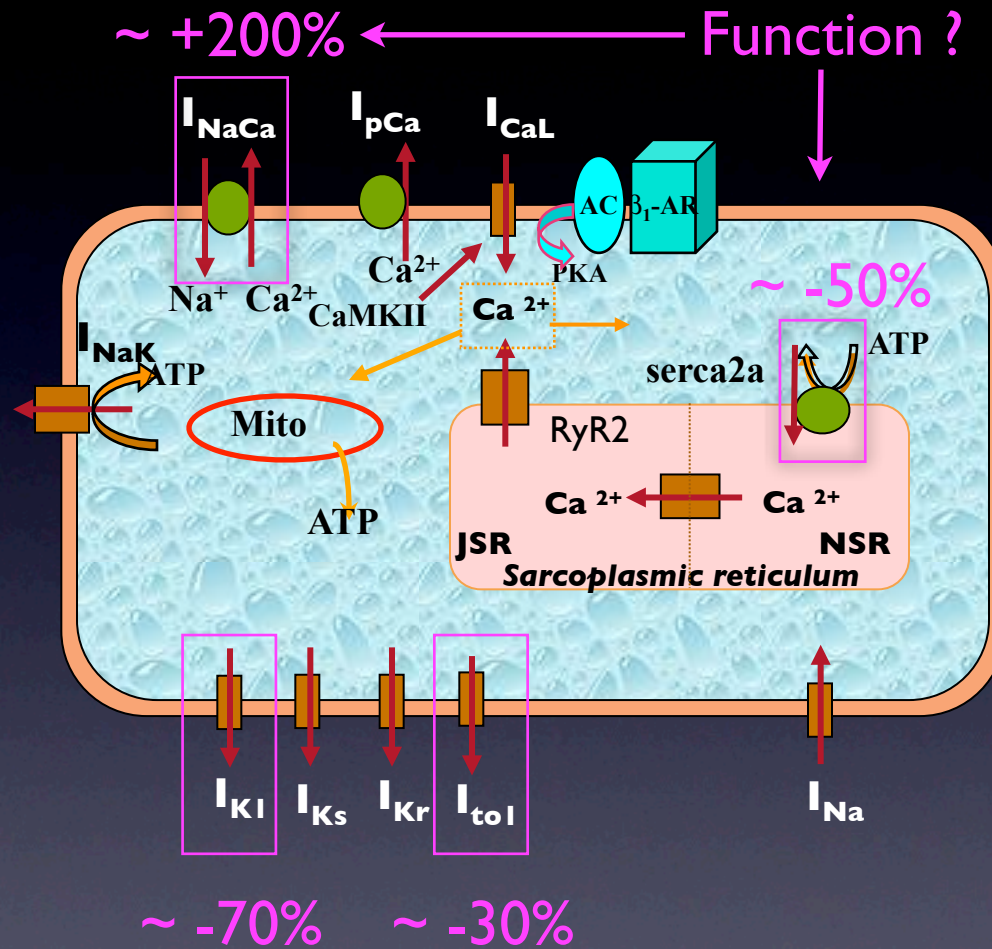
Hypothesis: Reduced Outward Current Prolongs APD

# The Molecular Phenotype of Heart Failure



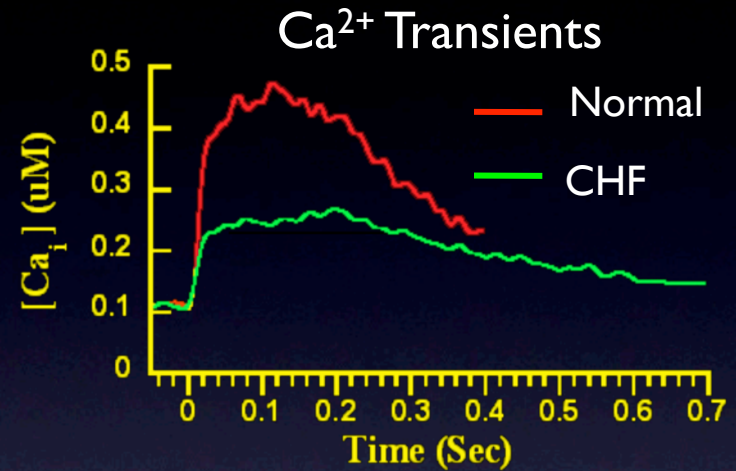
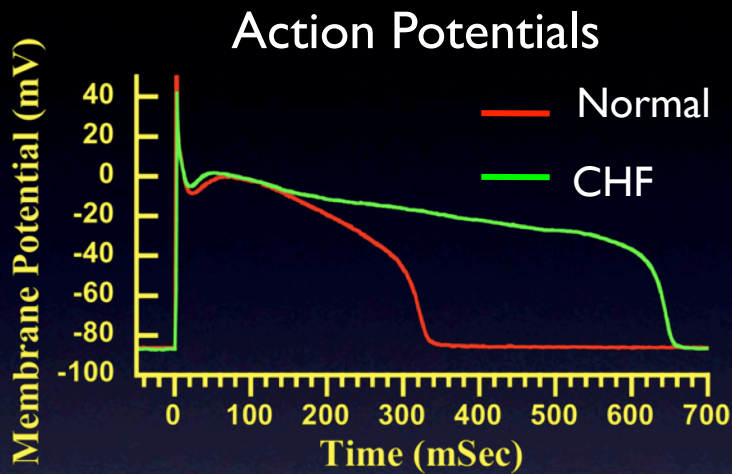
Hypothesis: Reduced Outward Current Prolongs APD

# The Molecular Phenotype of Heart Failure



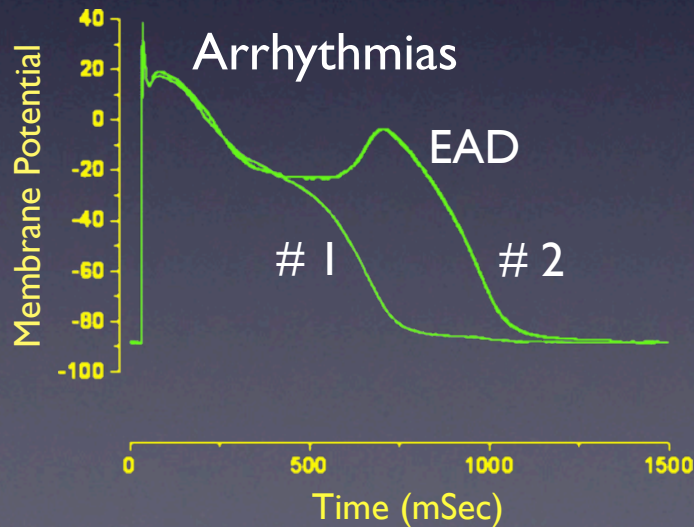
Hypothesis: Reduced Outward Current Prolongs APD

# The Cellular Phenotype of Heart Failure



O'Rourke, et al. Circ. Res. 84: 562, 1999

Winslow et al. Circ. Res. 84: 571, 1999

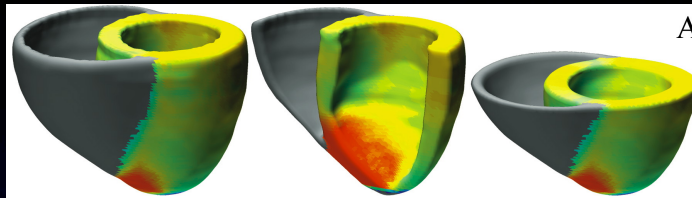


- APD prolongation
- reduced Ca<sup>2+</sup> transient magnitude
- Increased APD increases risk of EADs

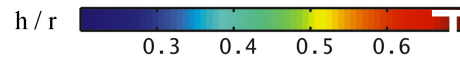
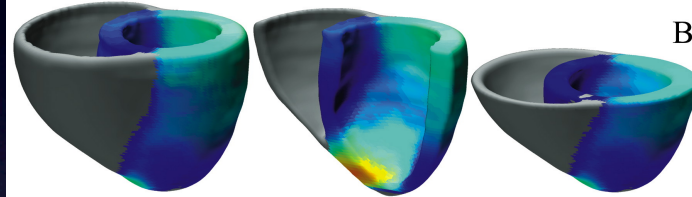
# The Anatomic Phenotype of Heart Failure

Helm et al (2006) Circ. Res. 98(1): 125

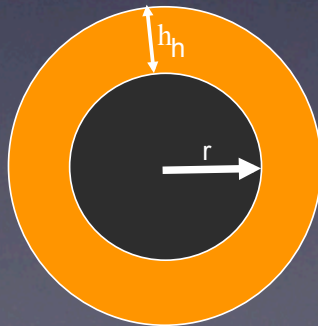
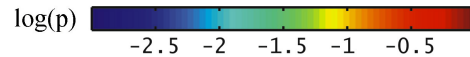
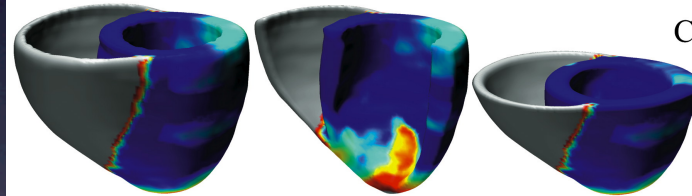
RWT  
Normal



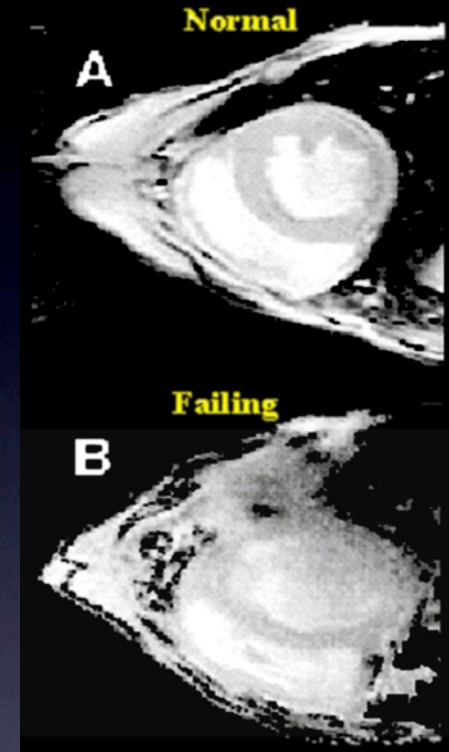
RWT  
Failing



log(p)



Relative Wall Thickness (RWT) =  $h/r$




MR heart image pre- and post-tachycardia pacing (McVeigh et al)

# Modeling Approaches

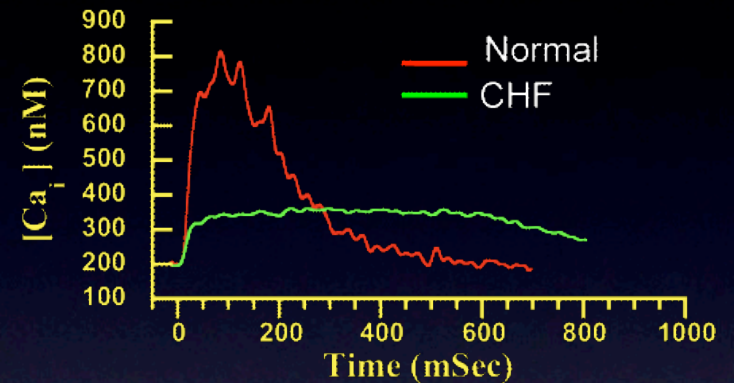
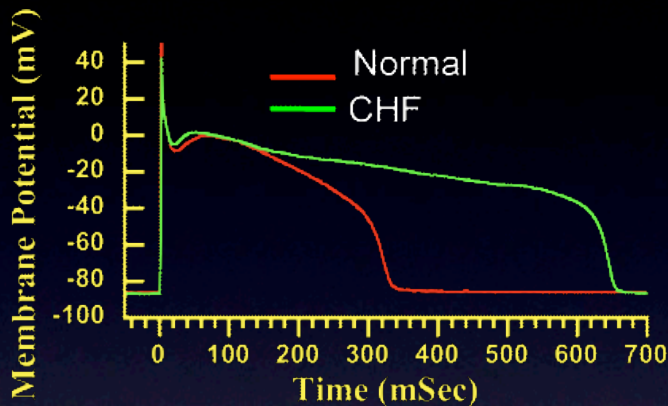
- *“Nano-scale” model of  $Ca^{2+}$  signaling in the dyad*
  - Fokker-Planck equation
- *“Meso-scale” models of integrative cell function*
  - Stochastic ordinary differential equations
  - Deterministic ordinary differential equations
- *“Macro-scale” models of ventricular conduction*
  - reaction-diffusion partial differential equations
- *There are approaches for moving across these scales*

# Dis-regulation of Protein Expression and Cell Function

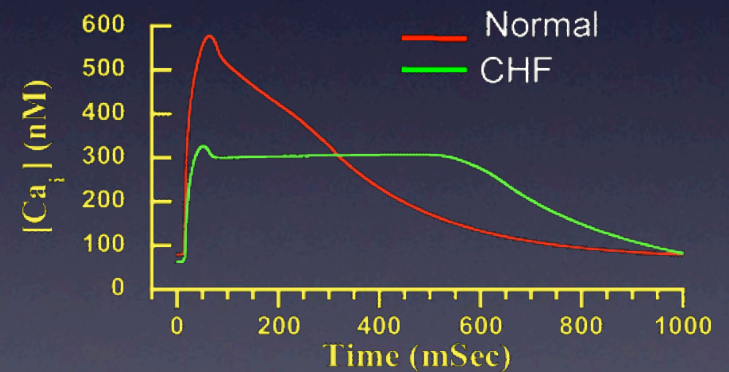
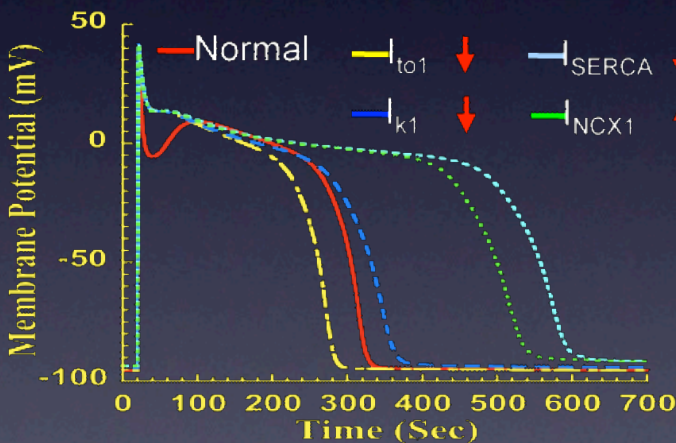
- ***Molecular phenotype***
    - altered protein expression
    - disruption of protein co-localization
  - ***Pathway phenotype***
    - increased B-adrenergic input, PKA hyper-phosphorylation
  - ***Cellular phenotype***
    - Action potential prolongation, repolarization abnormalities
  - ***Organ phenotype***
    - Reduced contractility
    - High risk of re-entrant arrhythmias
- 

# Relate Molecular to Cellular Phenotype Using “Meso-Scale” ODE Model

Experiments

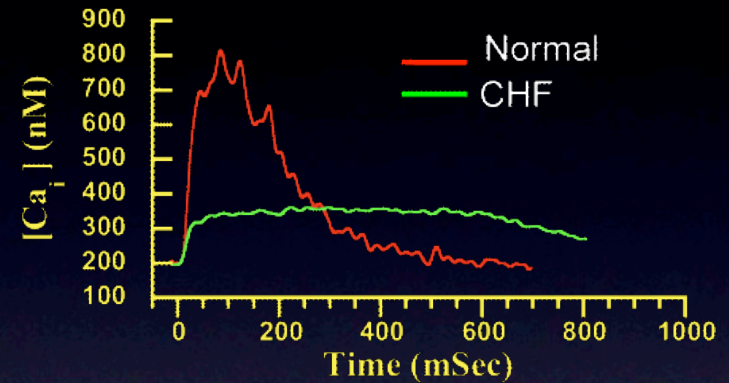
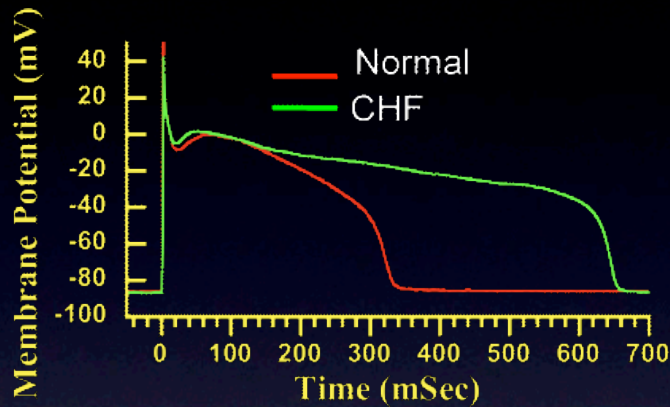


Models

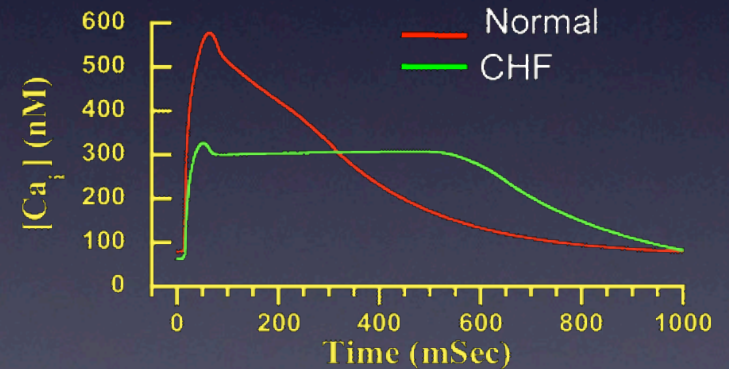
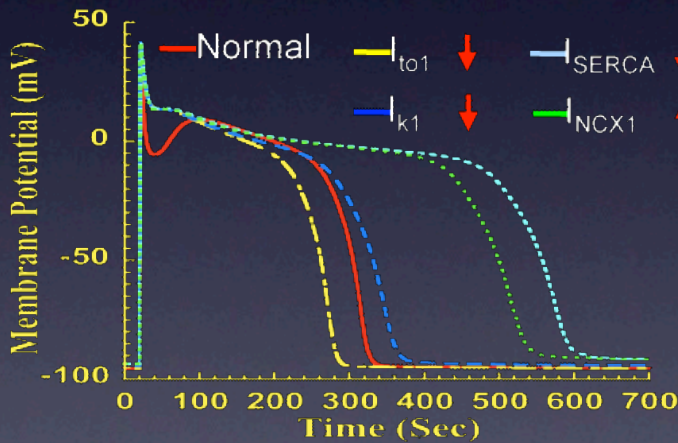


# Relate Molecular to Cellular Phenotype Using “Meso-Scale” ODE Model

Experiments



Models

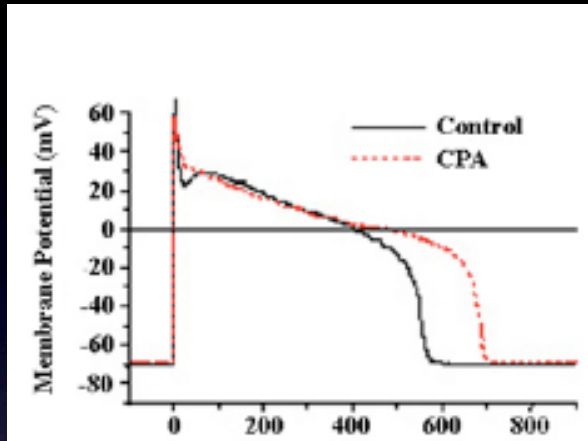


Non-Intuitive Result: Altered  $Ca^{2+}$  Handling Key to APD Prolongation

# Experimental Validation of Model Predictions

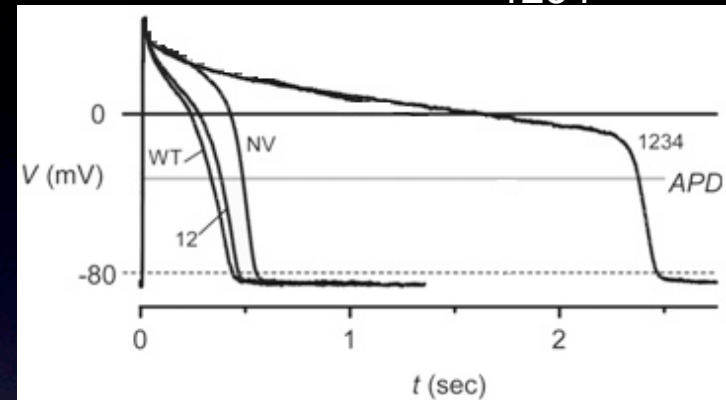
Experiment

## Serca Reduction



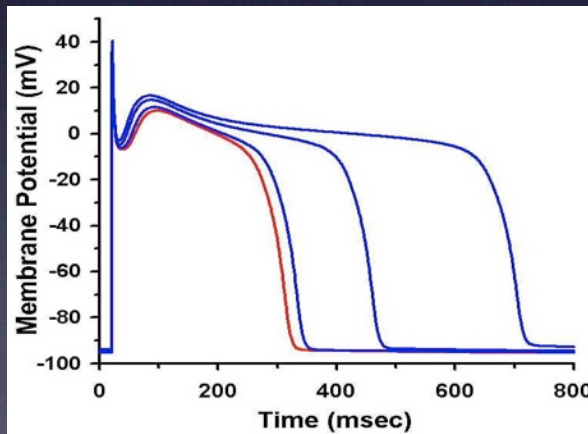
Winslow et al *Phil. Trans. Roy. Soc. Lond. A* 359: 1187

## Mutant CaM<sub>1234</sub>

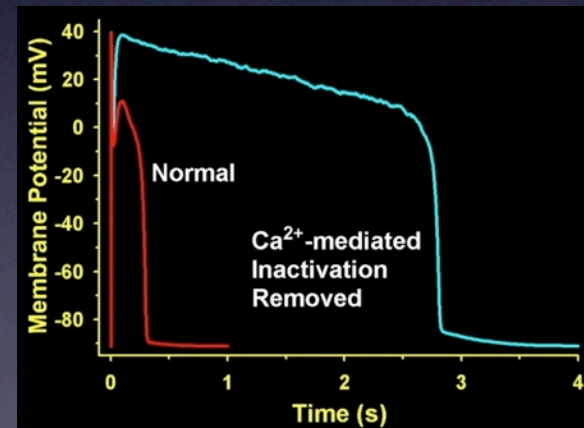


Alseikhan et al *PNAS* 99(26): 17185

Model



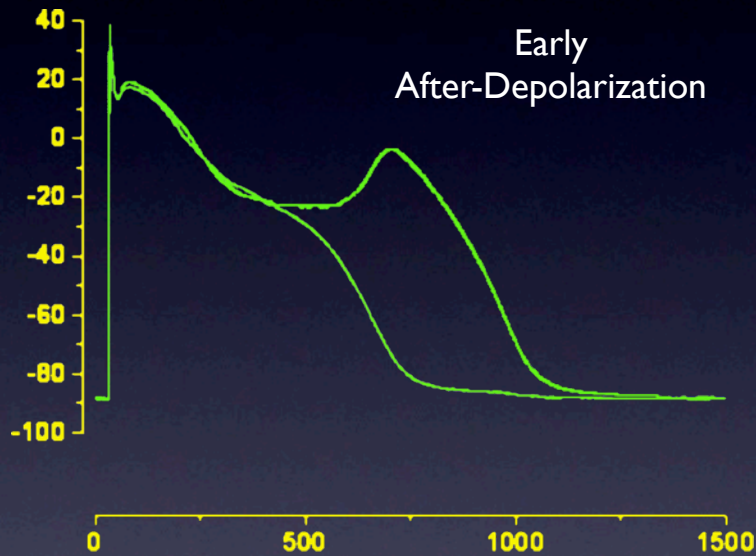
Winslow et al *Phil. Trans. Roy. Soc. Lond. A* 359: 1187



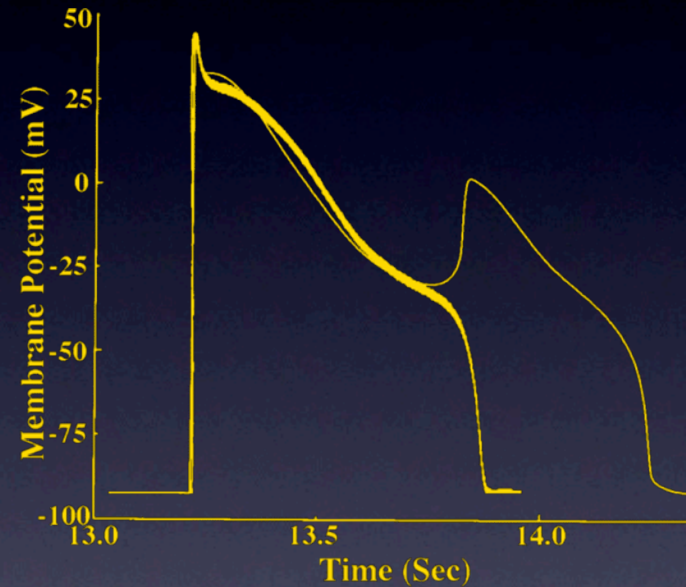
Greenstein & Winslow et al *Biophys. J.* 83(6): 2918

# Functional Consequences of APD Prolongation in HF

Experiment




Model



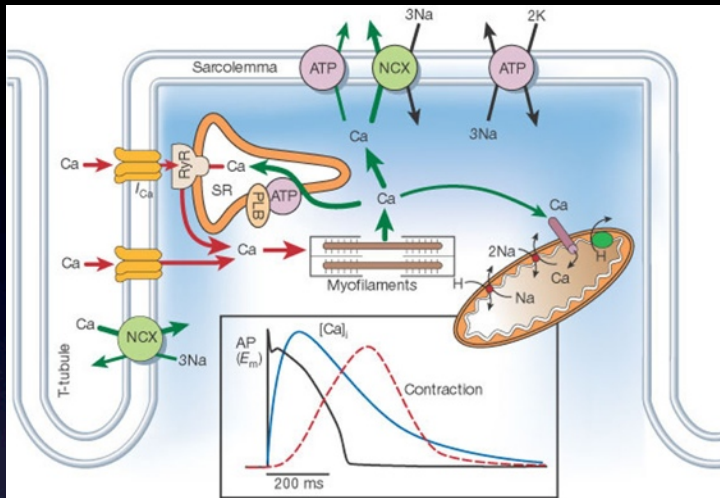
APD Prolongation  $\Rightarrow$  EADs

Models Have Explained the Mechanism

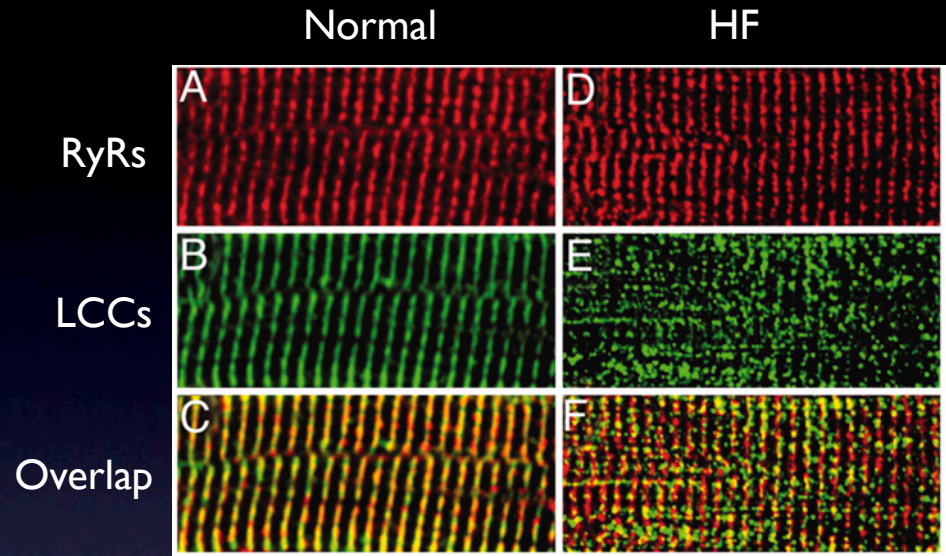
# Protein Localization and Mechanical Function

- ***Molecular phenotype***
    - altered protein expression
    - disruption of protein co-localization
  - ***Pathway phenotype***
    - increased B-adrenergic input, PKA hyper-phosphorylation
  - ***Cellular phenotype***
    - Action potential prolongation, repolarization abnormalities
  - ***Organ phenotype***
    - Reduced contractility
    - High risk of re-entrant arrhythmias
- 

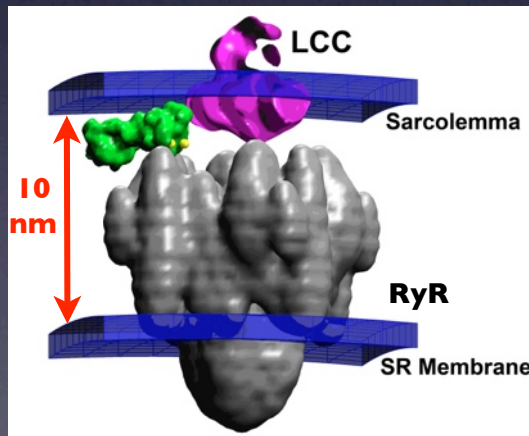
# The Architecture of Ca<sup>2+</sup> Signaling



Bers (2002) Nature 415(6868): 198



Song et al (2006). PNAS 103(11): 4305



Structure of the LCC (4Å), calmodulin (1Å) and RyR (14Å)

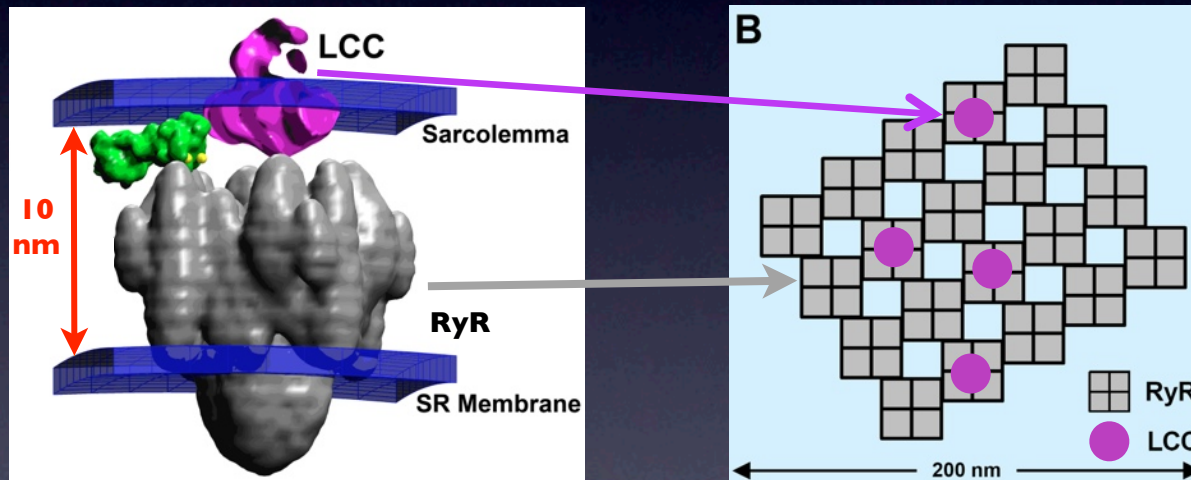
Number of Ca<sup>2+</sup> ions mediating LCC-RyR signaling may be very small

LCC & RyR co-localization may be disrupted

Functional consequences ?

# “Nano-scale” Model of $\text{Ca}^{2+}$ Signaling in the Dyad

Model dyad    LCC:RyR 4:20    ~5,000 dyads/cell




# “Nano-scale” Model of $\text{Ca}^{2+}$ Signaling in the Dyad

## Brownian Motion in a Potential Field

$$\frac{\partial P(\mathbf{r}_1, \dots, \mathbf{r}_N, t)}{\partial t} = D \sum_{i=1}^N \frac{\partial}{\partial \mathbf{r}_i} \cdot \left[ \frac{1}{k_B T} \frac{\partial V(\mathbf{r}_1, \dots, \mathbf{r}_N)}{\partial \mathbf{r}_i} P(\mathbf{r}_1, \dots, \mathbf{r}_N, t) + \frac{\partial P(\mathbf{r}_1, \dots, \mathbf{r}_N, t)}{\partial \mathbf{r}_i} \right] \quad \text{FP Eq.}$$

$$\frac{\partial}{\partial \mathbf{r}_i} = \left[ \begin{array}{ccc} \frac{\partial}{\partial r_{i,1}} & \frac{\partial}{\partial r_{i,2}} & \frac{\partial}{\partial r_{i,3}} \end{array} \right]$$

$$V(\mathbf{r}_1, \dots, \mathbf{r}_N) = \sum_{i=1}^N [2q\phi(r_i) + u(r_i)] + U(\mathbf{r}_1, \dots, \mathbf{r}_N)$$

  
Potential  
Energy

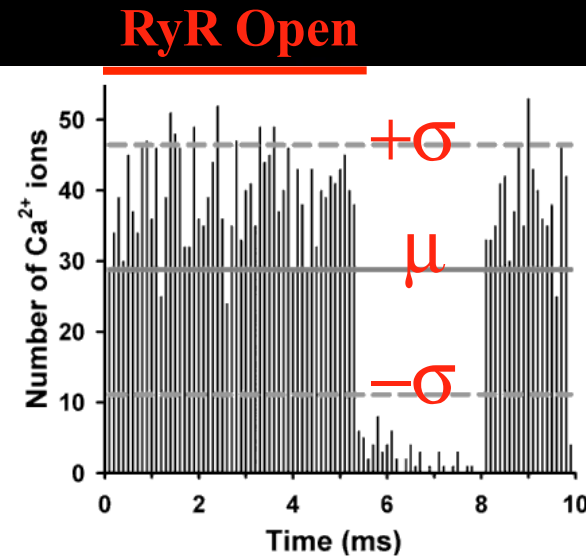
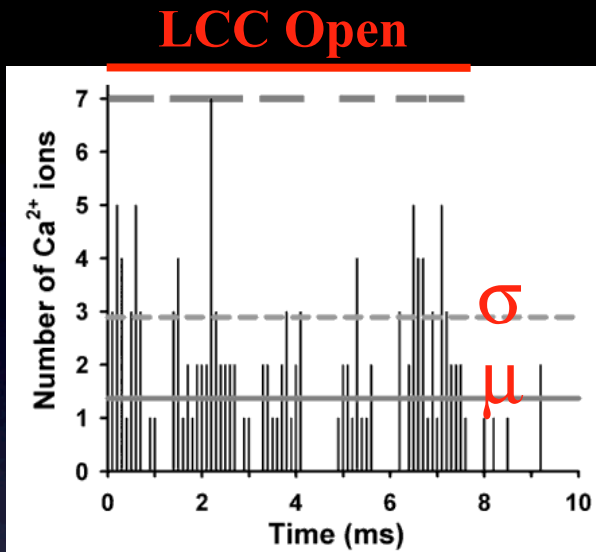
  
Surface  
Charge

  
Hard  
Core

  
Ion  
Interaction

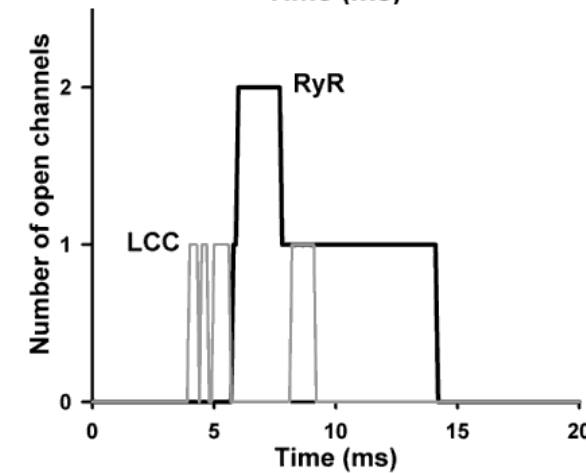
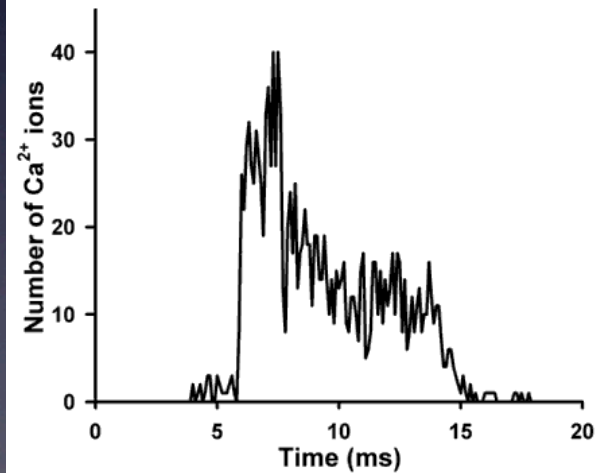
# Signaling Mediated by Ten's of $\text{Ca}^{2+}$ Ions

Single LCC  
 0 mV  
 Open at  $t=0$   
  
 $1.4 \pm 1.4$   
 $\text{Ca}^{2+}$  ions



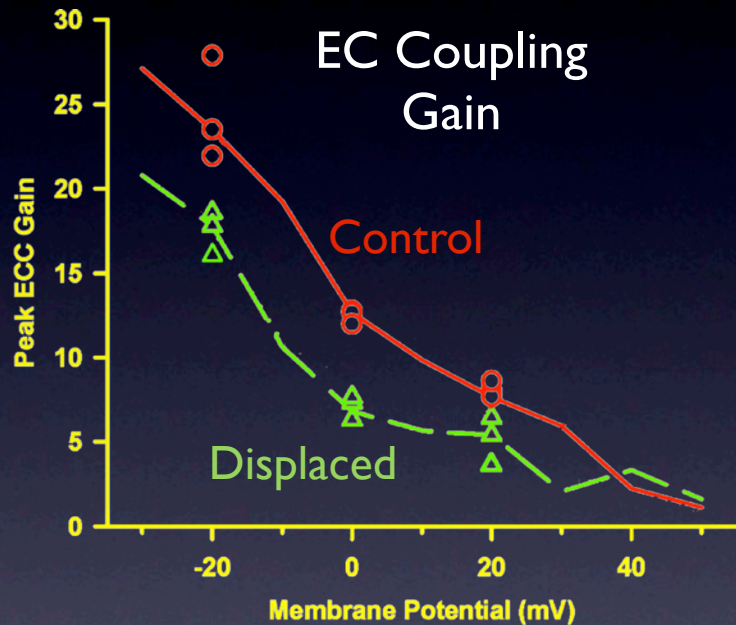
Single RyR  
 0 mV  
 Open at  $t=0$   
  
 $28.8 \pm 17.7$   
 $\text{Ca}^{2+}$  ions

Single CaRU  
 4 LCC, 20 RyR2  
 0 mV  
  
 $\sim 10$   $\text{Ca}^{2+}$  ions  
 In late phase



Tanskanen et al Biophys. J. 92: 3379

# Disruption of LCC-RyR Co-Location



ECC gain very *sensitive* function of relative channel placement

Reduced gain => less Ca<sup>2+</sup> release

Disruption in heart failure could impair muscle contraction

$$\text{EC Coupling Gain} = \frac{\text{Total Ca}^{2+} \text{ Release Flux}}{\text{Total Ca}^{2+} \text{ Trigger Flux}}$$

# Altered Signaling and Cellular Arrhythmias

- ***Molecular phenotype***

- altered protein expression
- disruption of protein co-localization

- ***Pathway phenotype***

- increased B-adrenergic input, PKA hyper-phosphorylation

- ***Cellular phenotype***

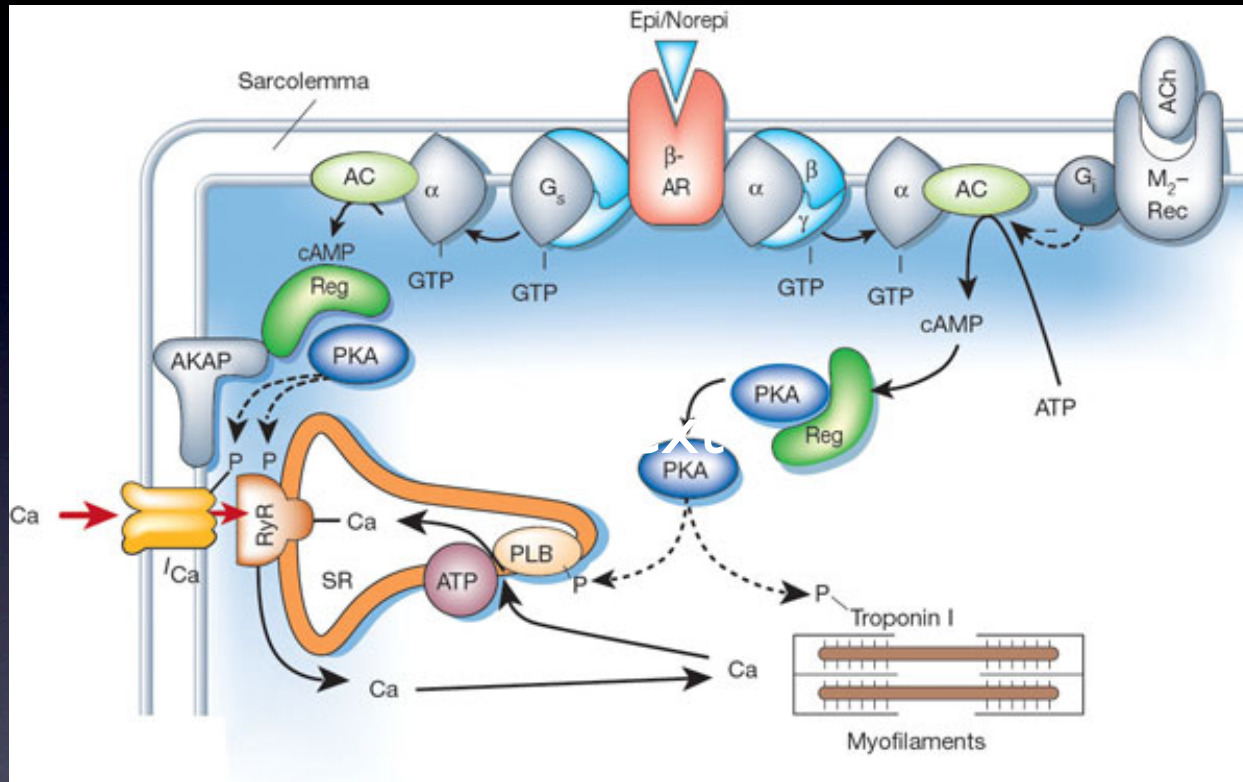
- Action potential prolongation, repolarization abnormalities

- ***Organ phenotype***

- Reduced contractility
- High risk of re-entrant arrhythmias



# Hyper-Phosphorylation of LCCs in HF

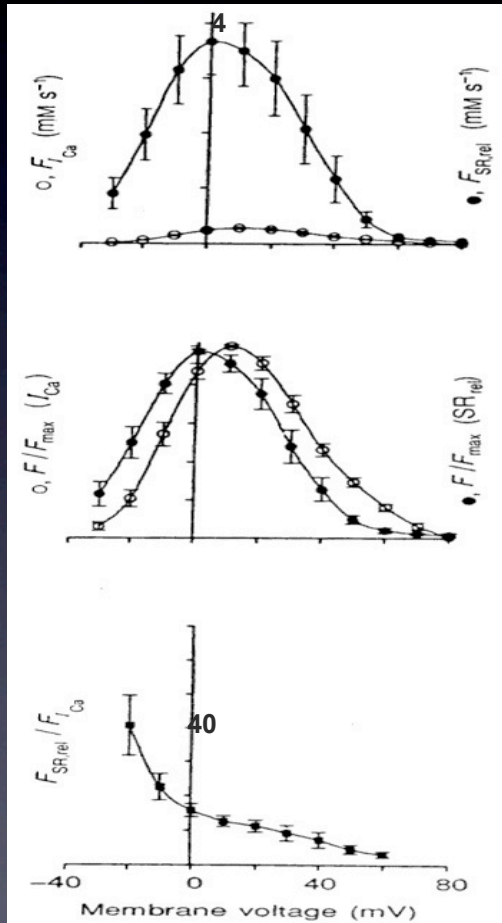


Phosphorylated LCCs have 10x longer mean open times

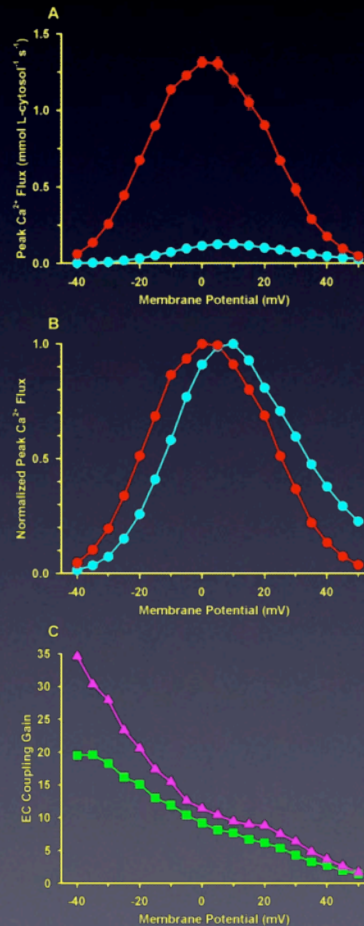


# Stochastic Model Reproduces Important Properties of $\text{Ca}^{2+}$ Release

## Experiment



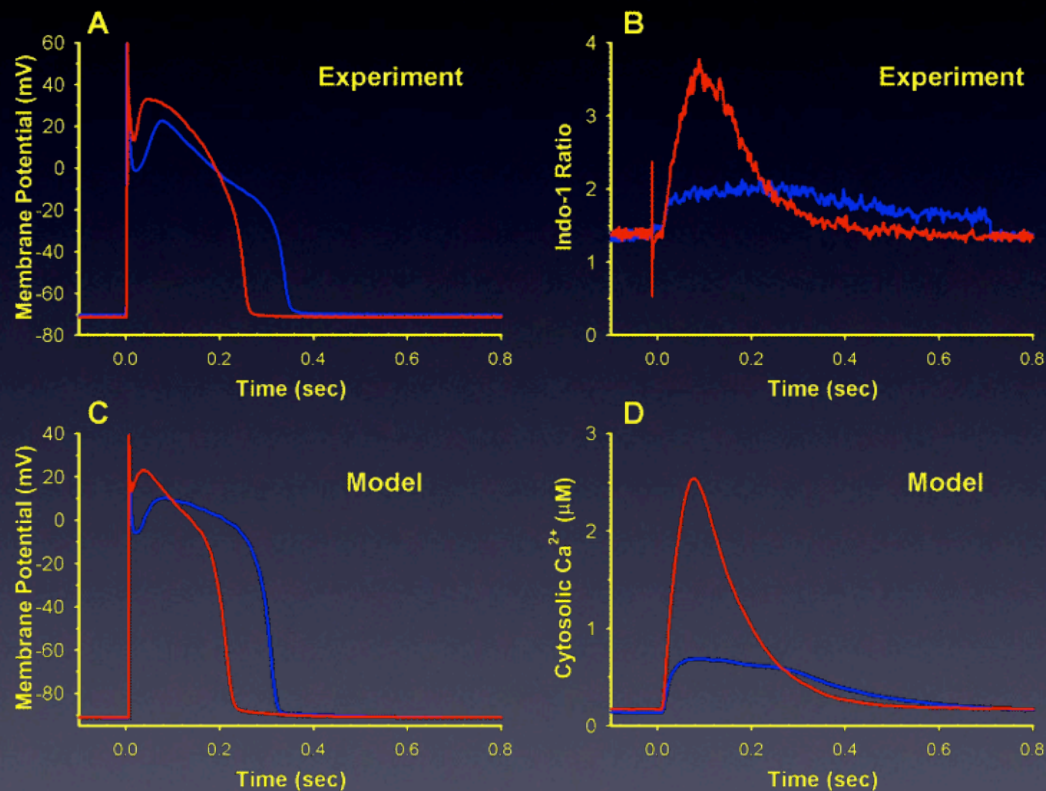
## Model



*~ 10 Sec for 1 simulation-  
Sec on 64 dual-core Intel  
2.8 GHz, MHz nodes*

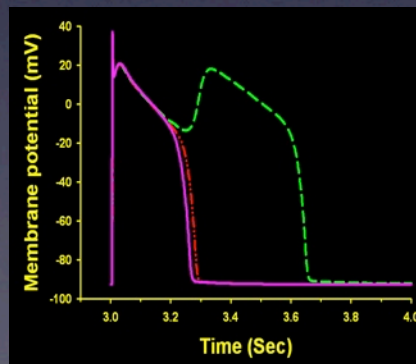
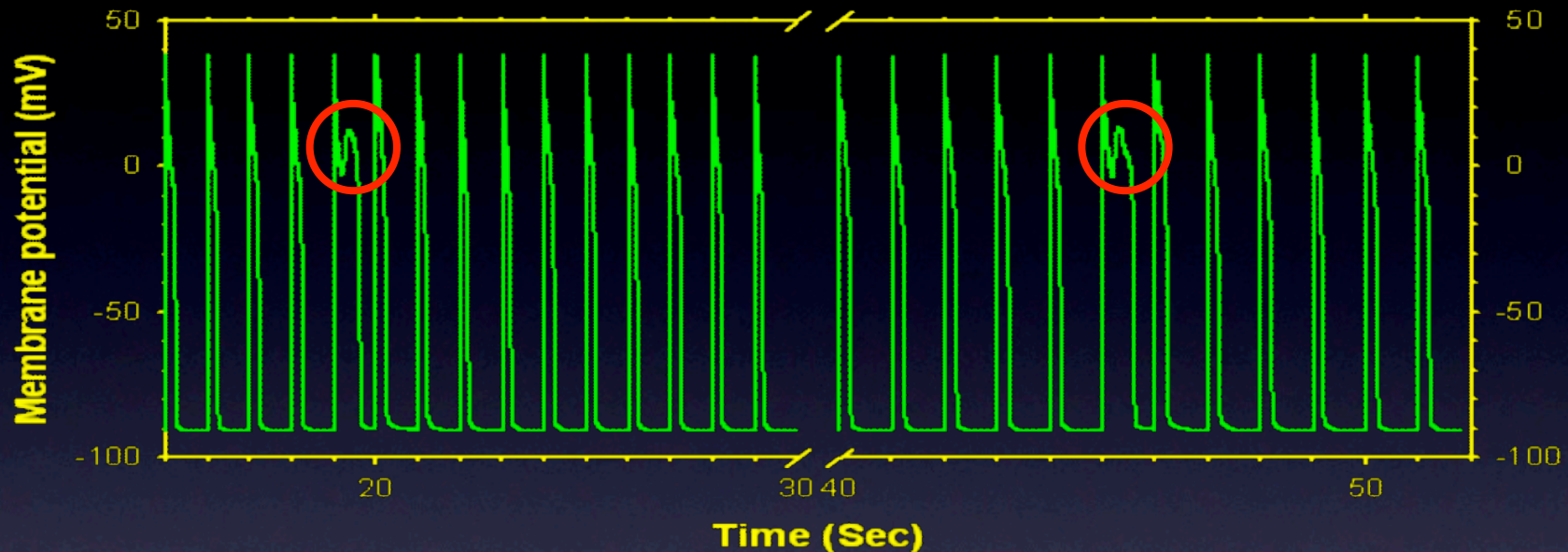
Greenstein & Winslow, Biophys. J. 83: 2918-2945

# Use Stochastic Cell Model to Study Chronic PKA Activation in HF



- **$\beta$ -AR activation of PKA**
  - Chronic activation in HF
  - phosphorylates PLN, up-regulates serca2 $\alpha$  pump
  - phosphorylates and down-regulates membrane  $K^+$  current
  - phosphorylates LCCs, switch from Mode 1 to Mode 2 gating
  - mean channel open time in Mode 2 is 10x larger than in Mode 1

# Functional Consequence of Increased Open Time: Random Early After-Depolarizations



- EADs produced by PKA phosphorylation of  $\text{Ca}^{2+}$  channels
- MECHANISM - Clustered openings of  $\text{Ca}^{2+}$  channels with long open time
- EADs inherently stochastic, discovered using stochastic ODE model
- EADs correlated with Sudden Cardiac Death

# Cellular to Whole-Heart Arrhythmias

- **Molecule**

- dis-regulation of protein expression
- disruption of protein co-localization

- **Pathway**

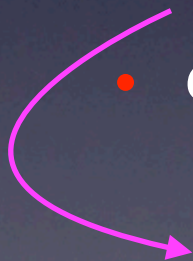
- PKA pathway & hyper-phosphorylation

- **Cell**

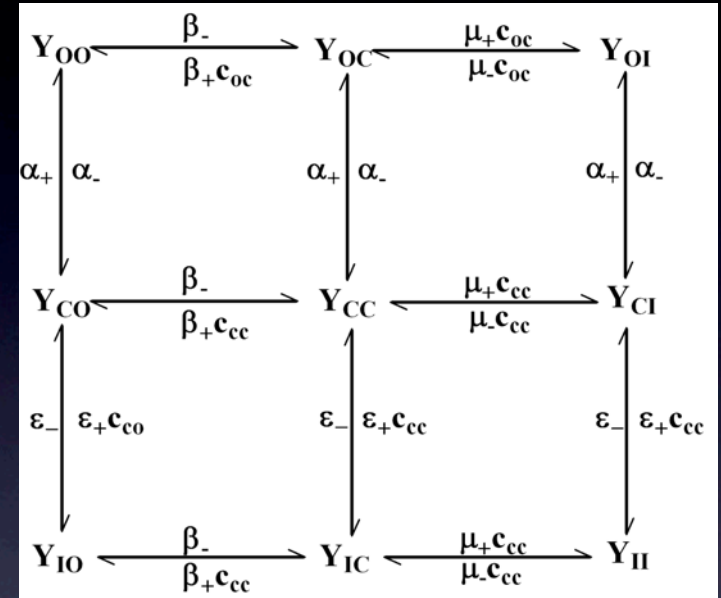
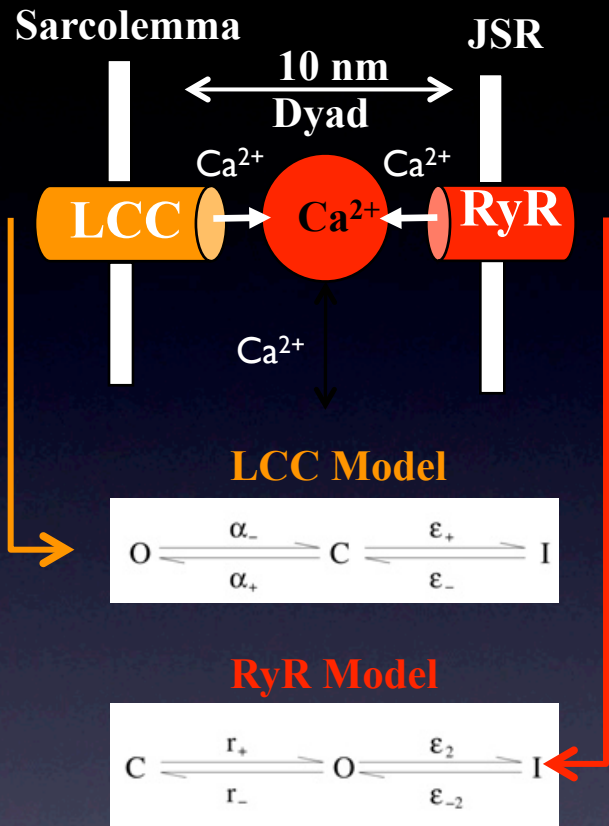
- Action potential prolongation, cellular arrhythmias

- **Organ**

- Reduced contractility
- Re-entrant arrhythmias



# Step from the Stochastic ODE Model to ODE Model Using the Rapid Equilibrium Approximation

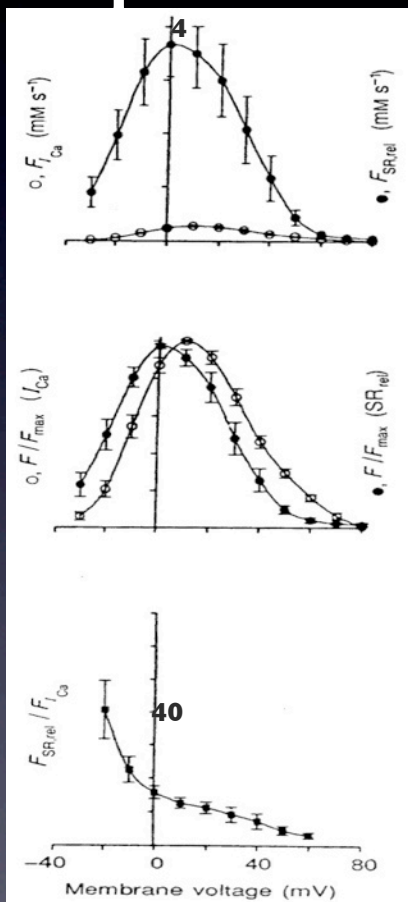


Hinch et al Biophys. J. 87: 3723

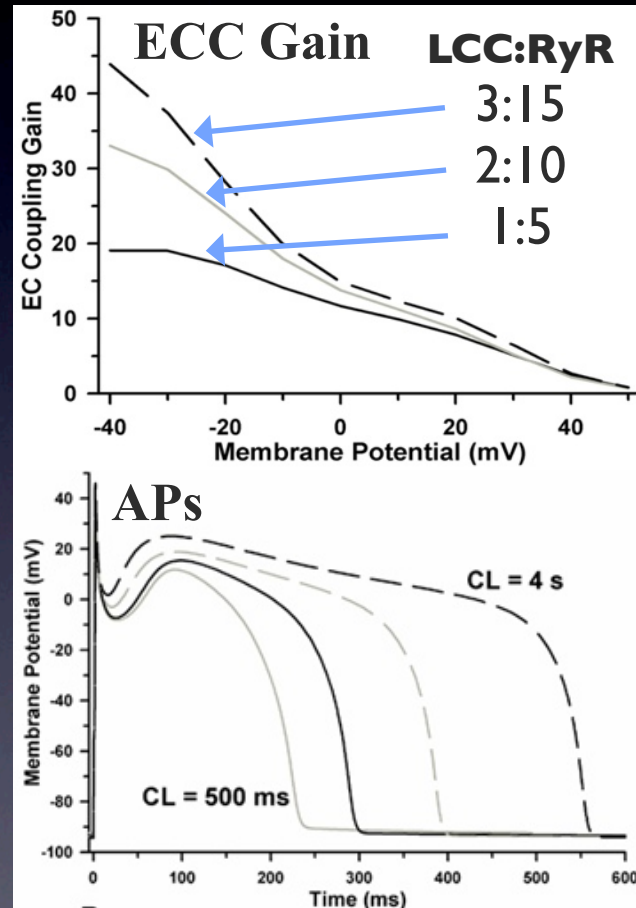
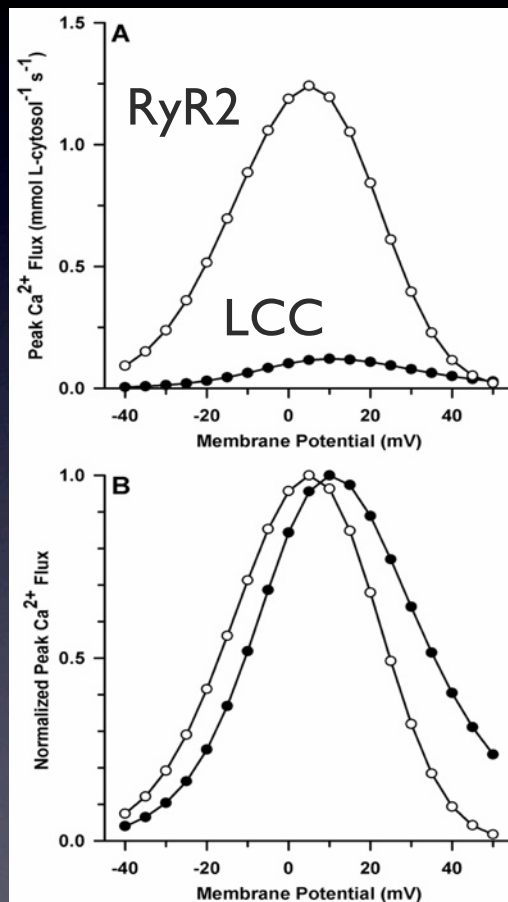
- Dyad  $Ca^{2+}$  changes fast ( $\sim 1 \mu\text{Sec}$ ) wrt channel kinetics ( $\sim 100$ 's  $\mu\text{Secs}$ )
- Dyad  $Ca^{2+}$  is in rapid equilibrium, algebraic function of other slow state variables
- Complex model reduced to low-dimensional system of coupled ODEs
- $\sim 10 \text{ mSec}$  for 1 simulation-Sec on one dual-core Intel Xeon 2.8 GHz node

# Resulting ODE Model Reconstructs Broad Range of Data, Can be Used in Whole Heart Simulations

## Experiment

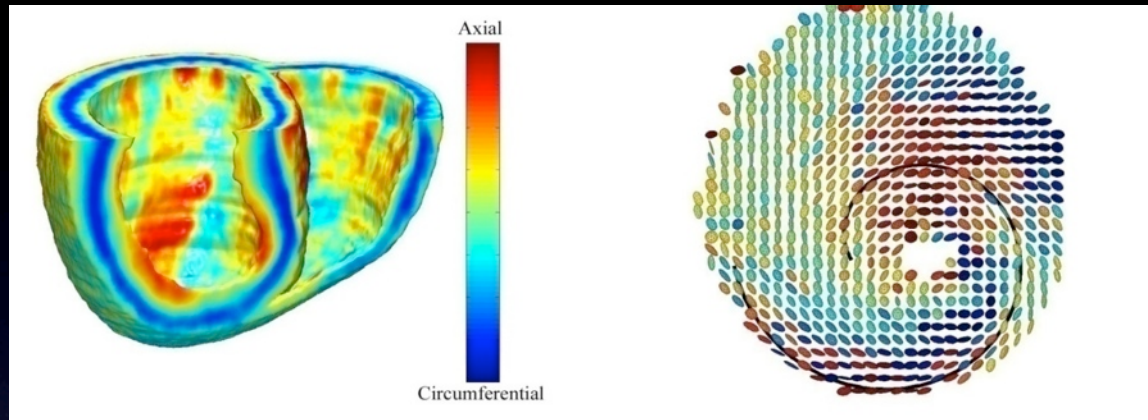


## Model

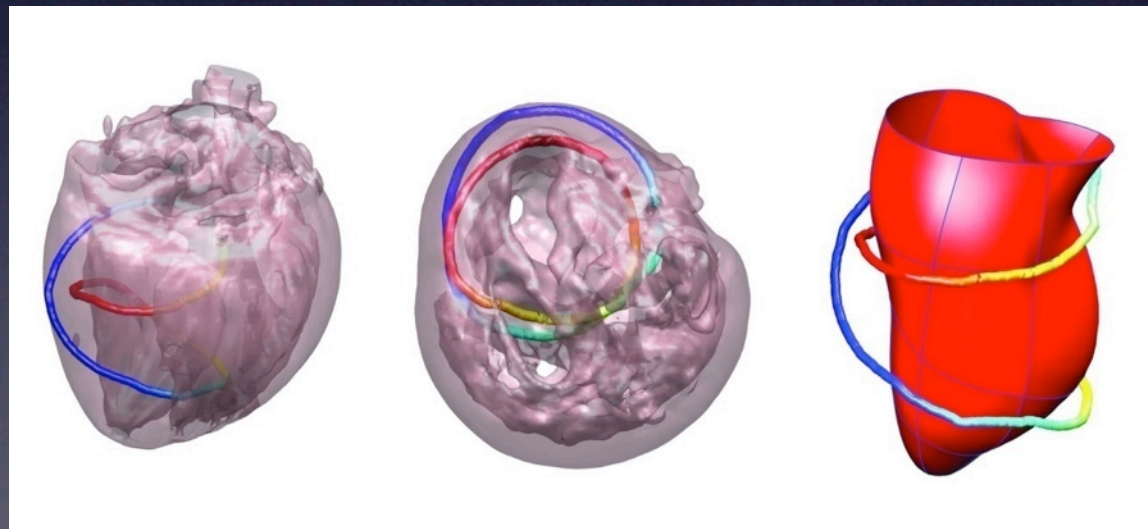
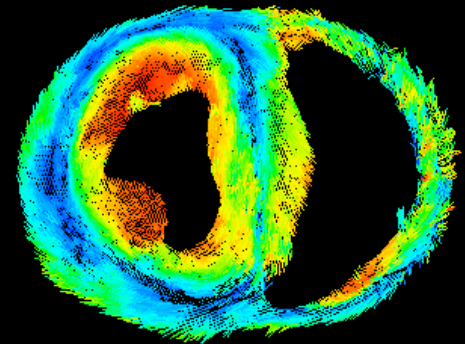


# Diffusion Tensor MR Imaging

## Whole-Heart Geometry, Fiber & Sheet Structure



Helm, PA (2005). *Magn. Reson. Med.* 54(4): 850



Helm, PA (2005). *Circ. Res.* 98(1): 125

- Fixed Heart (canine)
- 3D-FSE
- Imaging time ~ 60 Hrs
- 1.5T Clinical Magnet
- 350  $\mu\text{m}$  in-plane, 800  $\mu\text{m}$  out-of-plane resolution
- $> 10^6$  myocardial points

# “Macro-scale” Model of Whole-Heart

$$\frac{\partial v(\underline{x}, t)}{dt} = \frac{1}{C_m} \left[ \underbrace{-I_{ion}(v(\underline{x}, t))}_{\text{Reaction Term}} - \underbrace{I_{app}(\underline{x}, t)}_{\text{Applied Current}} + \frac{1}{\beta} \frac{\kappa}{1 + \kappa} \nabla \cdot (M_i(\underline{x}) \nabla v(\underline{x}, t)) \right] \quad \forall \underline{x} \in H$$

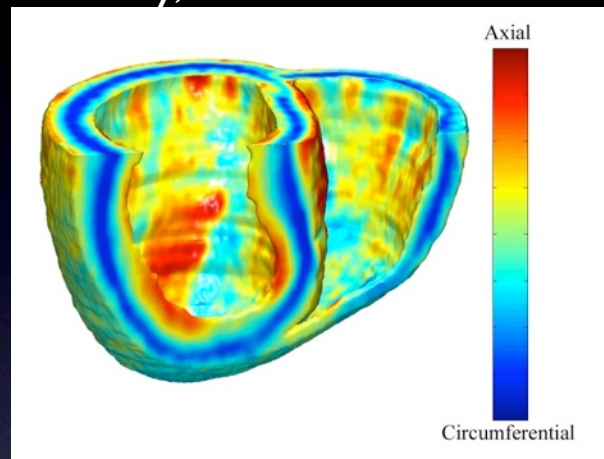
- **Model describes electrical conduction in the cardiac ventricles**
- **Constraining data**
  - ***H*** and **diffusion term** defined by heart fiber structure
  - **Reaction term**  $I_{ion}(\underline{x}, t)$  defined by the whole cell model state variables
- **Model structure**
  - Mesh myocardium at 100 um resolution,  $\sim 10^6$  mesh elements
  - Numerically integrate resulting system of coupled ODEs ( $\sim N \times 10^6$  ODEs)
  - *1 Simulation-Sec in  $\sim 3.5$  hrs on 64 nodes*
  - $\sim 8 * N$  GB/simulation-sec of data, where N is # state variables

# Model Validation

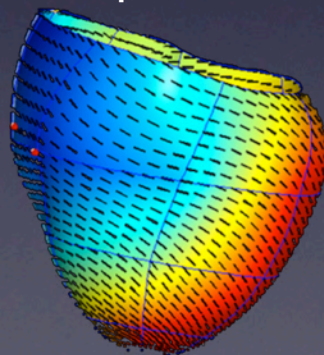
## Whole Heart Electrical Mapping



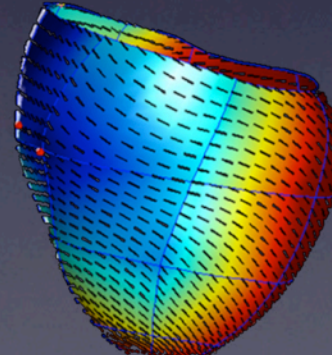
## Imaging-Based Measurement of Heart Geometry, Fiber and Sheet Structure



## Experiment



## Model

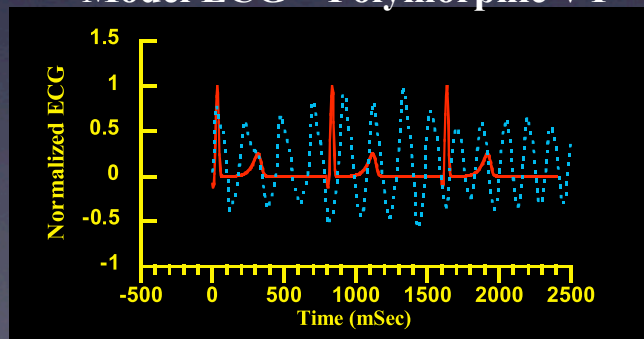


# Use “Macro-Scale” Model to Study Mechanisms of Arrhythmia in HF

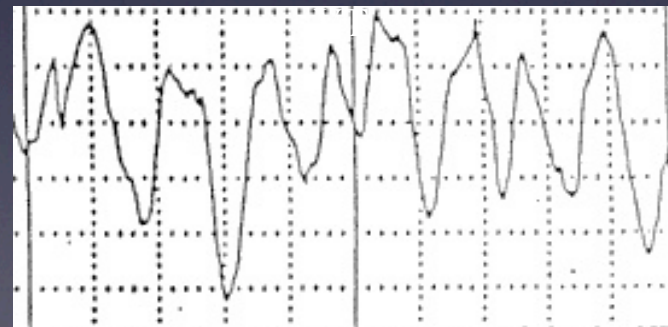
**Simulation: Normal Heart Conduction  
2 Stimulus Pulses**

**Simulation: Failing Heart, Single Stimulus  
Evokes EAD**

**Model ECG – Polymorphic VT**



**Experimental ECG – Polymorphic**

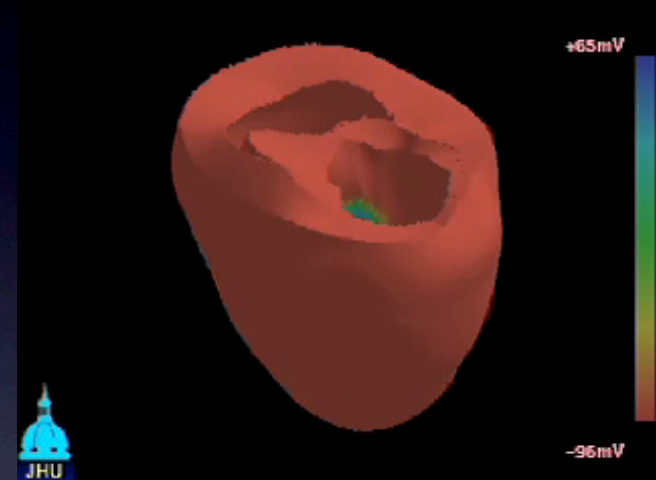


# Use “Macro-Scale” Model to Study Mechanisms of Arrhythmia in HF

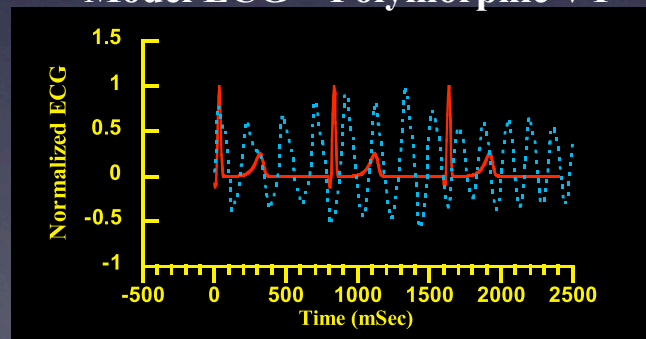
Simulation: Normal Heart Conduction  
2 Stimulus Pulses



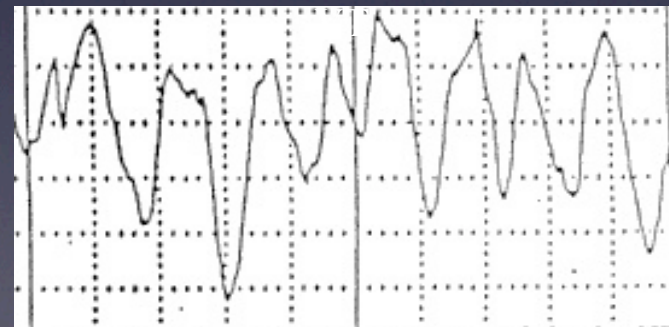
Simulation: Failing Heart, Single Stimulus  
Evokes EAD



Model ECG – Polymorphic VT

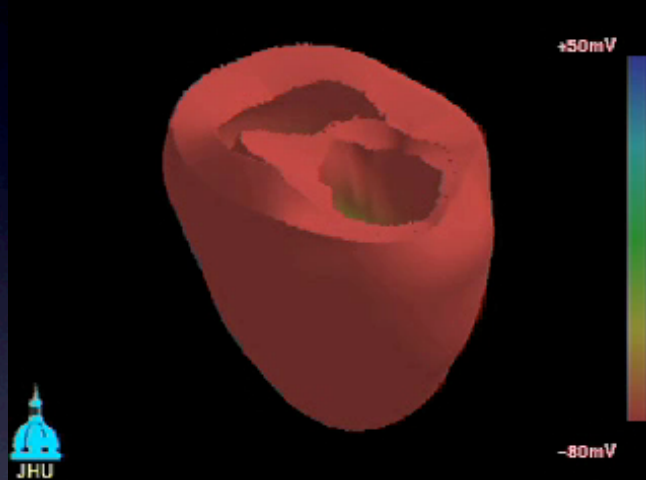


Experimental ECG – Polymorphic

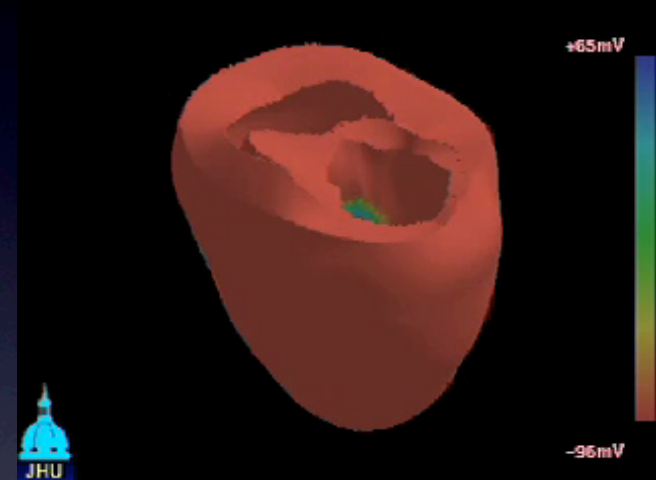


# Use “Macro-Scale” Model to Study Mechanisms of Arrhythmia in HF

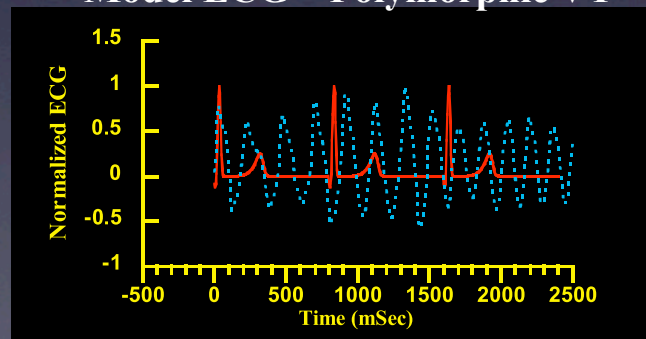
Simulation: Normal Heart Conduction  
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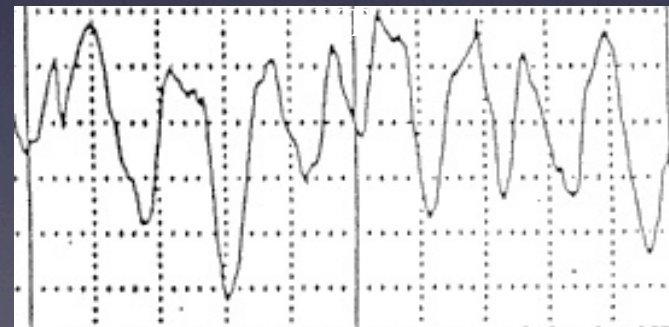
Simulation: Failing Heart, Single Stimulus  
Evokes EAD



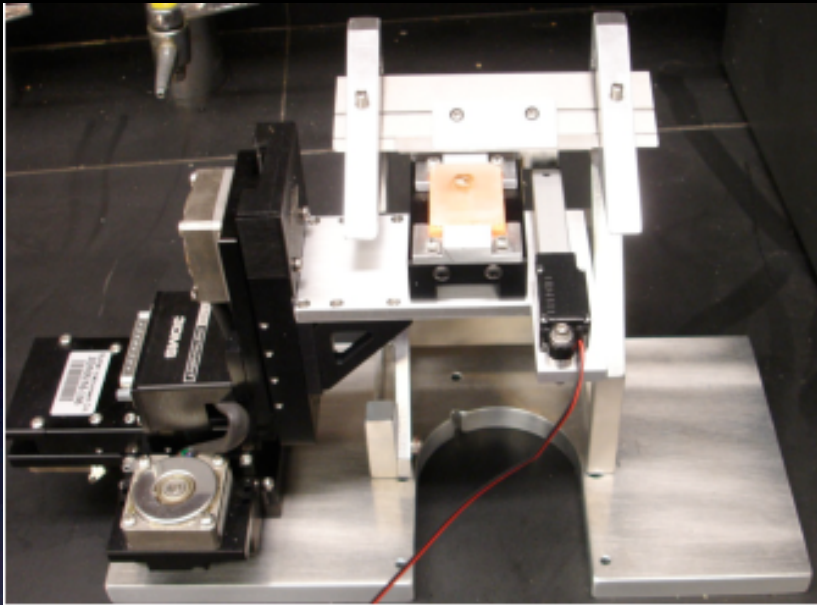
Model ECG – Polymorphic VT



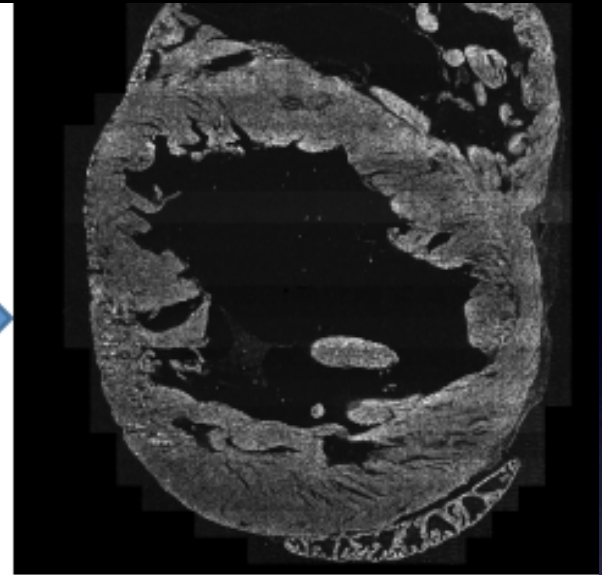
Experimental ECG – Polymorphic



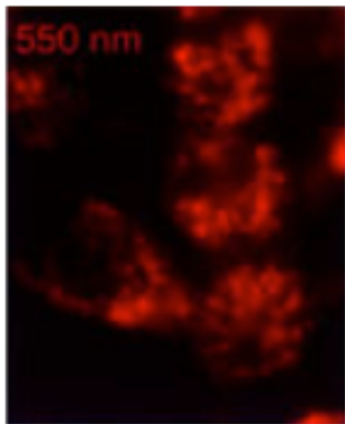
# Next Step: Super-High Resolution ( $\sim 500$ nm) Anatomic Reconstruction of Whole Hearts for Modeling



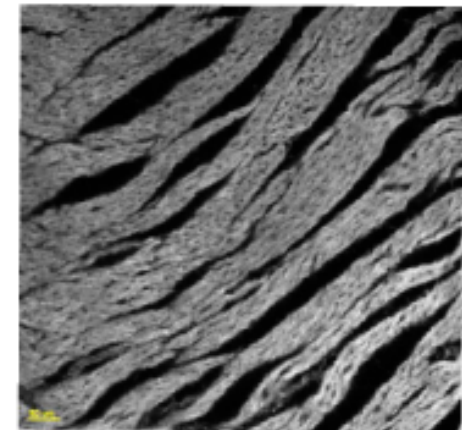
Raster Scan  
of Surface



Mitochondria



Cells



# Challenges

- *Local signaling between proteins in micro-domains is a common biological design motif*
  - *protein structure, charge, position, hard calculations*
- *Regulation of cellular electrical function by gene and protein-signaling networks*
  - *few biological studies, need quantitative data, expands the complexity of models*
- *“Macro-scale” models and arrhythmia generation*
  - *Difficult to constrain and validate models, computational complexity*

# Acknowledgements

## Myocyte Modeling

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**Saleet Jafri**  
**Reza Mazhari**  
**Jeremy Rice**  
**Sean Sun**  
**Antti Tanskanen**  
**An-Chi Wei**

## Tissue & Heart Modeling

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**Patrick Helm**  
**Alex Holmes**  
**David Scollan**

## Experiments

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**McVeigh Lab**  
**O'Rourke Lab**  
**Tomaselli Lab**  
**Yue Lab**

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